

# CHEMICAL CONSTITUENTS AND RADICAL SCAVENGING ACTIVITY OF METHANOL LEAF EXTRACT OF Vitex doniana



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Abstract:	This study was aimed at investigating the chemical constituents and radical scavenging activity of methanol leaf extract of <i>Vitex doniana</i> . The 2, 2 diphenyl-1-picrylhydrazyl (DPPH) radical scavenging activity, trace metal contents and phytochemical constituents of <i>V. doniana</i> leaf extract were carried out. The results demonstrated that the leaf extract possess a high radical scavenging activity with an IC <sub>50</sub> value of 198.53 µg/ml and 139.52 µg/ml for quercetin which was used as standard. Forty three phytochemicals were identified in the leaf extract of <i>V. doniana</i> , major among them include linalool (7.89%), β-Bisabolene (7.22%), erucic acid (7.11%), D-Nerolidol (6.22%), α-Farnesene (5.57%), α-Caryophyllene (5.28%), Calarene (1.02%), α-Copaene (0.80%). The metal element composition showed that the leaf extract is rich in some trace elements such as Mn, Fe and Zn. The presence of the phytochemicals and trace metals (iron, zinc, selenium, manganese and copper) identified in the leaf extract of <i>V. doniana</i> could be responsible for its high 2,2-diphenyl-1-picrylhydrazyl radical scavenging activity.
Keywords:	Leaf extract, phytochemicals, radical scavenging, Vitex doniana

# Introduction

The use of plants as medicine is as old as the existence of man on earth. Plants are known to possess secondary metabolites with pharmacological activities. Most of these phytochemicals are antioxidants (Lee et al., 2017). Vegetables, fruits and other edible plants are vital dietary sources of vitamins and essential metals. (Zhai et al., 2015). Medicinal plants have been explored and utilized as herbal medicine in the treatment of several diseases (Petrovska, 2012). Owing to intensified research, it is been reported that a lot of plants actually possess medicinal worth (Sofowora, 1993). The use of nutritional antioxidants in combating pathological disorders related to oxidative stress has attracted attention in recent years. Products of plant origin are known to exert their ameliorative effects by cleaning up free radicals and adjusting antioxidant defense to the system (Tarasub et al., 2011). Plants of tropical and sub-tropical climate are known to possess large amount of phytochemical antioxidant compounds as secondary metabolites which is a natural adaptive way of surviving in a harsh environments. In recent decades, evidences abound from epidemiological and laboratory studies to show that some edible plants possess great potential in the prevention, management and remedy of a wide range of human diseases (Agbafor and Nwachukwu, 2011).

Vitex doniana (V. doniana) of the Verbanaceae family is a perenial shrub with wide distribution in tropical West Africa such as Nigeria, and extends to some East African countries and in the savannah and high rainfall areas (Atawodi et al., 2003). Vitex doniana is a deciduous tree, usually 4-8m high, occasionally up to 15m, with a heavy rounded crown (Burkill, 2000). V. doniana is an edible plant. In the middle belt of Nigeria, like Kogi state, the tender leaves are utilized as vegetables for sauces and porridge for meals (Yakubu et al., 2013). V. doniana leaves is consumed in Nigeria as vegetables and valued sources of protective foods that is of great benefits for the maintenance of good health and disease prevention (Nnamani et al., 2007). In ethno-medicine, V. doniana leaf is utilized in the management and treatment of many diseases. It is employed in the treatment of stomach and rheumatic pains, inflammatory disorders, diarrhoea and dysentery (Iwueke et al., 2006). According to Iwu (1993), the leaves and roots of V. doniana are also utilized in the treatment of nausea, colic and epilepsy. Vitex doniana is widely used in traditional system of medicine. The fruit of V. doniana is sweet and tastes like prunes; it is at times sold and eaten as food. It contains vitamins A and B and can be made into a jam. The leaves are commonly used as herb and for food (Katende, 1995). The

fruit can be prepared into wine, and pounded leaves can also be added to warm filtered grain beer and then drunk. The bark yields a dye that can be used for cloth (Katende, 1995).

*V. doniana* has been utilized as medication for liver disease, stiffness, leprosy, backache, hemiplegia, conjunctivitis, rash, measles, rachitis, febrifuge, as tonic galactagogue to aid milk production in nursing mothers, sedative, digestive regulator and treatment of eye problems, kidney problems and as supplement for lack of vitamin A and B (Burkill, 2000). This study is therefore aimed at investigating the chemical constituents and radical scavenging activity of the methanolic leaf extract of *V. doniana* 

# Materials and Methods

## Collection and extraction of Vitex doniana leaves

Fresh leaves of *Vitex doniana* were collected within the premises of Kogi State University Anyigba, Nigeria. They were thereafter confirmed in the Department of Biological Sciences, Kogi State University, Anyigba, Nigeria. The plant leaves were rinsed in distilled water, air dried and pulverized. To obtain the methanol leaf extract of *Vitex doniana* (MEVd), 1000 g of the plant material were extracted in 3 litres of methanol for 48 h. The filteration process was done using Whatman grade 1 filter paper and vacuum pump. The extract was concentrated in vacuum at 40°C with a rotary evaporator (Model Modulyo 4K, Edward England), and water bath to dryness. The percentage (%) yield of the methanol leaf extract of *Vitex doniana* was 8.25% and the crude extracts obtained was stored in capped vials at 4°C.

## DPPH radical scavenging activity assay

The stable 2, 2 diphenyl-1-picrylhydrazyl (DPPH) radical was used for the determination of free radical scavenging activity of the plant extract. The method as modified by Arogba and Omede (2012) was used for the determination of free radical scavenging activity of the plant extract. To a portion (1 ml) each of the different concentrations of the extract or standard (Quercetin) in test tubes was added 1 ml of 0.3 mM DPPH in methanol. The mixture was vortexed and then incubated in a dark chamber for 30 min after which the absorbance were read at 517 nm against a DPPH control containing only 1 ml of methanol in place of the extract. The experiment was carried out in triplicate and the percentage scavenging activity was calculated using the expression:

% Scavenging Activity =  $[(Ac - As)/Ac] \times 100$ 

**Where:** Ac = Absorbance of control; As = Absorbance of sample

#### Identification of the chemical constituents of the extract

Gas chromatography-mass spectrometry (GC-MS) analysis was conducted using GCMS -QP 2010 Plus Shimadzu Japan with a GC 2010 injector temperature of 250°C, column oven temperature of 80°C and carrier gas pressure of 108.0 kPa. The column length was 30 m with a diameter of 0.25 mm. The elutes were automatically passed into a mass spectrometer. The GC-MS-QP2010 Plus ion source temperature was 230°C; interface temperature of 250°C. Comparison of the GC-MS spectrograph of the sample with the instrument's data bank together with computer matching with the database of National Institute of Standard Technology (NIST) library revealed that the methanol leaf extract of V. doniana contained various bioactive compounds that eluted at different retention times depending on the boiling point of the eluted constituent. The plant extract sample was injected by splitting with the split ratio 1:60.

### Determination of trace metal contents of the extract

The metal content of the leaf extract was determined using atomic absorption spectrometry (AAS). The plant extract (1 g) was weighed into a digestion flask and 20 ml of acid mixture (650 ml conc. HNO3; 80 ml Perchloric acid (PCA); 20 ml conc. H<sub>2</sub>SO<sub>4</sub>) was added. The flask was heated until a clear digest was obtained. The digest was diluted with distilled water to the 500ml mark, and aliquots of the diluted clear digest were used for AAS using filters that match the different elements. The method was optimized based on the proper wavelengths for each element: Fe (248.3 nm), Mn (279.5 nm), Zn (213.9 nm), Se (196.0 nm), Cu (324.754 nm), Cr (357.869 nm), Pb (283.754 nm), Cd (228.802 nm), As (189.0 nm) and Hg (253.7 nm).

#### **Results and Discussion**

## Free radical scavenging activity of the leaf extract

The methanol leaf extract of V. doniana showed a very high  $(94.03 \pm 0.44\%)$  DPPH radical scavenging activity which is slightly higher than that of Quercetin (92.97  $\pm$  0.49) at the same concentration (1000 µg/ml). Also at lower concentration, the extract exhibited higher radical scavenging activity than Quercetin. However, the result of this study revealed that quercetin has a lower IC<sub>50</sub> value (139.52 $\mu$ g/ml) compared to the methanol extract of V. doniana leaves which has an IC<sub>50</sub> value of 198.53 µg/ml (Table 1). Free radicals have become popular in biological research as a result of their pivotal role in physiological processes and in addition to their involvement in various pathological disorders. Free radicals affect several significant biomolecules, and as a result disturb the normal redox balance leading to elevated oxidative stress (Phaniendra et al., 2015). This study revealed that the methanol leaf extract of Vitex doniana (MEVd) exhibited a high DPPH radical scavenging activity with an IC50 of 198.53 µg/ml which is, however, not as high as that of quercetin (139.52 µg/ml). This is an indication that MEVd may be an efficient free radicals scavenger. This is in agreement with the work of Agbafor and Nwachukwu (2011) which revealed that the leaf extracts of Vitex doniana possess antioxidant property.

 Table 1: DPPH Radical Scavenging Activity (%) of the methanol leaf extract of V. doniana

Concentration	% Radical scavenging.		
Concentration	Extract	Quercetin	
1000	$94.03 \pm 0.44$	$92.97 \pm 0.49$	
500	$87.62 \pm 0.92$	$85.29 \pm 0.80$	
250	$60.61 \pm 1.10$	$74.28 \pm 0.49$	
125	$51.53 \pm 0.96$	$64.85 \pm 0.47$	
62.50	$36.50 \pm 1.22$	$38.03 \pm 1.09$	
31.25	$22.52 \pm 1.14$	$15.86\pm0.86$	
$IC_{50}$ (µg/ml)	$198.53\pm0.96$	$139.52\pm0.70$	

 Table 2: Qualitative and quantitative assessment of some metals by AAS

Metal	Mn	Fe	Zn	Se	Cu
$ppm \times 10^{-3}$	95.3	92.1	64.2	35.6	21.5

#### Concentration of some trace elements in the leaf extract

The analysis on the methanol leaf extract of *V. doniana* for the presence and concentration of some trace elements is shown in Table 2. This result showed that the plant extract contained essential trace elements that may have probably contributed to the radical scavenging activity of the extract. The AAS analysis as carried out in this study showed that the methanol leaf extract of *Vitex doniana* contained essential metallic nutrients such as Iron (92.1 × 10<sup>-3</sup>), Zinc (64.2 × 10<sup>-3</sup>), Selenium (35.6 × 10<sup>-3</sup>) and Corper (21.5 × 10<sup>-3</sup>).

Zinc is a well recognized essential metal known to alleviate heavy metal toxicity. There are complex inter-relationships between cadmium and some essential trace elements. A number of essential trace elements such as zinc (Zn), iron (Fe), selenium (Se) and copper (Cu) play vital roles in controlling various metabolic and signaling pathways. Among the trace elements Zn and Fe are essential for maintenance of life and health. Zn is an essential trace metal with numerous functions in biological systems. It controls several enzymes of intermediary metabolism, DNA and RNA synthesis, gene expression, immune-competence and plays a significant role in homeostasis of hormones. Zn also takes part in the defense against excessive amounts and following damage of certain metals, and it does so through the interaction with metallothionein. It has been noted that Zn has a relationship with many enzymes in the body and can prevent cell damage through activation of the antioxidant defense system (Jamakala and Rani, 2014). Zn has similar chemical and physical characteristics with Cadmium. As a result it competes with Cadmium for the binding sites of metallic transporters and proteins of enzymatic function (Bridges and Zalups, 2005). Zn reduces the oxidative stress generated by Cadmium. This is as a result of the functional role as a cofactor of copper zinc-superoxide dismutase (Cu/Zn SOD) which is an antioxidant enzyme (Zhai et al., 2015). There are reports to show that Selenium (Se) intake provides protection against Cadmium toxicity in several organs including the liver and kidney. Se is a co-factor of glutathione peroxidase (GPx), the antioxidant enzyme which contributes to the antioxidant defense system. This capacity of Se empowers it to decrease Cadmium toxicity by decreasing Cadmium-induced oxidative stress and re-enforcing defense system of Cadmium-exposed host (Liu et al., 2013). Iron (Fe) competes with Cadmium for enterance into intestinal metal uptake transporters. This could explain the reduction in intestinal absorption of Cadmium following supplementation. The expression of these metal transporters is usually regulated by essential minerals such as Fe and Zn (Zhai et al., 2015). Manganese (Mn) is an essential trace mineral required in biological systems both as an activator and a constituent of several enzyme. Mn is a significant co-factor of mitochondrial superoxide dismutase, an antioxidant enzyme responsible for quenching and mopping out ROS (Eybl and Kotizová, 2010).

# Phytochemical constituents of the leaf extract

The GC-MS analysis showed that the methanol leaf extract of *V. doniana* comprised of forty three phytochemicals which eluted at various retention times based on the boiling point of the eluted component (Table 3). The biological activity, according to literature, of the major phytochemical constituents of the methanol leaf extract of *V. doniana* are shown in Table 4.

Table 3: Phytochemicals detect	ed in MEVd by GC-MS
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Peak No.	Name of Compound	R.T.	Molecular Formula	M.W	P.A. (%)
1	Acnetol acetate	4.026	$C_5H_8O_3$	116	0.76
2	β- <i>cis</i> -Ocimene	4.791	$C_{10}H_{16}$	136	0.39
3	Propane, 1,1,3-triethoxy-	5.118	$C_9H_{20}O_3$	176	0.42
4	Linalool	5.571	$C_{10}H_{18}O$	154	7.89
5	Phenylethyl Alcohol	5.900	$C_8H_{10}O$	122	1.02
6	Benzeneacetic acid, ethyl ester	7.503	$C_{10}H_{12}O_2$	164	0.95
7	Annulene	8.457	$C_8H_8$	104	3.67
8	Phenylmethanimine	9.084	$C_7H_7N$	105	3.55
9	Eugenol acetate	9.396	$C_{12}H_{14}O_3$	206	0.41
10	Lauryl chloride	9.534	$C_{12}H_{25}Cl$	204	1.52
11	Trans-a-Berganotene	9.969	$C_{15}H_{24}$	204	1.19
12	β- Bisabolene	10.144	$C_{15}H_{24}$	204	7.22
13	1H-Cyclopropa[a]naphthalene, 1a,2,3,5,6,7,7a,7b-octahydro-1,1,7,7a-tetramethyl-, 1aR- (1a.α.,7.alpha.,7a.α.,7b.α.)]-	10.306	$C_{15}H_{24}$	204	1.02
14	β–Farnesene	10.390	C15H24	204	1.46
15	α-Caryophyllene	10.605	C15H24	204	5.28
16	α –Farnesene	11.045	$C_{15}H_{24}$	204	5.57
17	α–Cubebene	11.355	C15H24	204	1.64
18	Copaene	11.486	C15H24	204	0.80
19	D-Nerolidol	11.772	$C_{15}H_{26}O$	222	6.22
20	$\alpha$ - Tetradecene (Neodene)	11.984	$C_{14}H_{28}$	196	0.77
21	Z,Z,Z-4,6,9-Nonadecatriene	12.210	$C_{19}H_{34}$	262	1.08
22	12-Oxabicyclo[9.1.0]dodeca-3,7-diene, 1,5,5,8-tetramethyl-, [1R-(1R*,3E,7E,11R*)]-	12.498	$C_{15}H_{24}O$	220	0.98
23	γ-Eudesmol	12.730	$C_{15}H_{26}O$	222	1.03
24	Eudesm-4(14)-en-11-ol	13.000	$C_{15}H_{26}O$	222	3.04
25	Hexadecyl mercaptan	14.412	$C_{16}H_{34}S$	258	0.65
26	Amyl ketone	15.265	$C_{11}H_{22}O$	170	3.19
27	Palmitic acid, ethyl ester	15.555	$C_{18}H_{36}O_2$	284	1.57
28	Methyl 14-methylpentadecanoate	16.728	$C_{17}H_{34}O_2$	270	1.73
29	Docosanoic acid, ethyl ester	17.371	$C_{24}H_{48}O_2$	368	0.54
30	Stearic acid, ethyl ester	18.095	$C_{20}H_{40}O_2$	312	1.87
31	11-Octadecenoic acid, methyl ester	19.912	$C_{19}H_{36}O_2$	296	2.87
32	Erucic acid	20.971	$C_{22}H_{42}O_2$	338	7.11
33	4-O-decyl-d-glucitol	21.325	$C_{16}H_{34}O_{6}$	322	5.00
34	Oleic acid amide	21.478	C <sub>18</sub> H <sub>35</sub> NO	281	1.84
35	Z-11-Hexadecenoic acid	22.170	$C_{16}H_{30}O_2$	254	1.18
36	Olealdehyde	22.649	C <sub>18</sub> H <sub>34</sub> O	266	0.68
37	Eicosane, 1-cyclohexyl-	22.979	$C_{26}H_{52}$	364	2.19
38	Phthalic acid, dioctyl ester	24.869	$C_{24}H_{38}O_4$	390	0.90
39	Stearic acid, ethyl ester	25.220	$C_{20}H_{40}O_2$	312	1.03
40	Behenic alcohol	25.534	$C_{22}H_{46}O$	326	0.99
41	Ethylamine, 2-((p-bromo . αmethyl αphenylbenzyl)oxy)-N,N-dimethyl-	25.927	$C_{16}H_{22}BrNO$	347	1.50
42	Ethylamine, 2-((p-bromo $\alpha$ methyl $\alpha$ phenylbenzyl)oxy)-N,N-dimethyl-	26.830	$C_{18}H_{22}Br$	347	3.40
43	Squalene	27.712	$C_{30}H_{30}$	410	3.91

## Table 4: Major phyto-components identified and their biological activities

PA (%)	Name of compound	MF	<b>Biological activity (according to literature)</b>
			Antioxidant (Seol et al., 2016), Anticancer (Sun et al., 2015), Anti-inflammatory (Maria et al., 2016);
7.89	Linalool	$C_{10}H_{18}O$	Anxiolytic (Cline et al., 2008); Antiseptic (Pengelly, 2004); sedative and local anesthetic effects
			(Cavanagh and Wilkinson, 2002); antinociceptic (Peana et al., 2004) activity.
0.39	β– <i>cis</i> –Ocimene		Antioxidant and anticancer (Mahmoud, 2013).
7.22	β- Bisabolene	$C_{15}H_{24}$	Anti-oxidant activity (Gholivand et al., 2010)
6.22	D-Nerolidol	$C_{15}H_{26}O$	Antioxidant (Vinholes et al., 2014); Antiulcer (Klopell et al., 2007); Antinociceptive (Koudou et al.
			2005); Antimalarial (Lopes <i>et al.</i> , 1999) activity.
5.57	α-Farnesene	$C_{15}H_{24}$	Antioxidant (Sarikurkcu et al., 2013); Anticarcinogenic (Afoulous et al., 2013); Antibacterial
			(Chehregani et al., 2010); Antifungal (Al-Maskri et al., 2011).
5.28	α-Caryophyllene	$C_{15}H_{24}$	Anti-inflammatory (Fernandes et al., 2007); Antitumour (Legault et al., 2003)
1.02	Calarene	$C_{15}H_{24}$	Antioxidant and antimicrobial activity (Wang <i>et al.</i> , 2010).
0.80	α – Copaene	$C_{15}H_{24}$	Antioxidant, anti – inflammatory and anticarcinogenic (Vinholes et al., 2013).
3.91	Squalene	$C_{30}H_{30}$	Anti-oxidant and Anticancer (GÜne§ (2013)
7.11	Erucic acid	$C_{22}H_{42}O_2$	Antioxidant, Analgesic and Anti-inflammatory (Ibrahim (2012).
1.02	Phenethyl alcohol	$C_8H_{10}O$	Antioxidant and antibacterial activities (Wang et al., 2015).

The GC-MS analysis of this study shows that linalool, a monoterpene, is the major component of the methanol leaf extract of *V. doniana*. A good number of recent studies have focused on the pharmacological activities of linalool. Linalool, a monoterpene alcohol, is a component of many natural aromatic plants. Linalool has been found to have

biological activities, including antioxidant (Seol *et al.*, 2016), anticancer (Sun *et al.*, 2015), anti-inflammatory (Wu *et al.*, 2014); anxiolytic (Cline *et al.*, 2008); Antiseptic (Pengelly, 2004); sedative and local anesthetic effects (Cavanagh and Wilkinson, 2002); antinociceptic (Peana *et al.*, 2004) activity. Sesquiterpenes are 15 – carbon compounds formed from 3

isoprenoid units and are secondary metabolites produced majorly in higher plants. It is one of the most familiar terpenes, are a class of natural products with a wide range of desired properties (Wang et al., 2011) Sesquiterpenes have been reported to exhibit antioxidant (Abolaji et al., 2013), anticarcinogenic (Afoulous et al., 2013), antimicrobial (Wang et al., 2013), antifungal (Kundu et al., 2013), antiinflammatory (Wang et al., 2013). In recent time, efforts are channeled into research and development of novel drugs from natural products. According to this study, most of the major bioactive components identified in the leaves of V. doniana are sesquiterpenes which include  $\beta$ -Bisabolene, D- Nerolidol, α- Farnesene, α-Caryophyllene, and calarene. β-Bisabolene have been reported to have biological activities, including anti-oxidant activity (Gholivand et al., 2010). D-Nerolidol, one of the major sesquiterpenes identified in the extract has been investigated to exhibit myriad of biological activities. There are reports to back-up the antioxidant (Vinholes et al., 2014); antiulcer (Klopell et al., 2007); cytotoxic (Sperotto et al., 2013); antinociceptive (Koudou et al. 2005), and antimalarial (Lopes et al., 1999) activity of Nerolidol. The sesquiterpene  $\alpha$ -Caryophyllene has been suggested to show a number of pharmacological and biological activities, including anti-inflammatory (Fernandes et al., 2007) and antitumour (Legault et al., 2003). a-Farnesene is found to be associated with many biological activities including antioxidant (Sarikurkcu et al., 2013), anticarcinogenic (Afoulous et al., 2013), antibacterial (Chehregani et al., 2010), and antifungal (Al-Maskri et al., 2011) properties. This study has shown that terpenes are the major compounds present in the fresh leaves of V. doniana. Owing to the fact that they are the main components, a number of the pharmacological activities and ethomedicinal uses of V. doniana can be attributed to the major terpenes present in the plant. Even so, the pharmacological and biological properties of V. doniana cannot be ascribed to only a single component, because the components present in the plant may interact in synergy to promote and/or re-enforce the antioxidant effect determined.

## Conclusion

From this study, it can be concluded that the methanol extract of *Vitex doniana* possessed a high 2,2-diphenyl-1picrylhydrazyl (DPPH) radical scavenging activity which could be as a result of the trace elements and phytochemicals identified in the leaf extract. These phytochemicals have been reported to have antioxidant, anti-inflammatory and anticancer activities.

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#### **Conflict of Interest**

The authors declare that they have no conflict of interests.

#### References

- Abolaji AO, Eteng MU, Omonua O & Adenrele Y 2013. Influence of coadministration of artemether and lumefantrine on selected plasma biochemical and erythrocyte oxidative stress induces in female wistar rats. *Hum. Exp. Toxicol.*, 32: 206-215.
- Afoulous S, Ferhout H, Raoelison EG, Valentin A, Moukarzel B, Couderc F, & Bouajila J. 2013. Chemical composition and anticancer, antiinflammatory, antioxidant and antimalarial activities of leaves essential oil of *Cedrelopsis grevei. Food Chem. Toxicol.*, 56: 352–362.

- Agbafor KN & Nwachukwu N 2011. Phytochemical analysis and antioxidant property of leaf extract of *Vitex doniana* and *Mucuna pruriens. Biochem. Res. Intl.*, 1: 1-4.
- Al-Maskri AY, Hanif MA, Al-Maskari MY, Abraham AS, Al-Sabahi JN & Al-Mantheri O 2011. Essential oil from Ocimumbasilicum (Omani Basil): A desert crop. Nat. Prod. Commun., 6: 1487–1490.
- Arogba SS & Omede A 2012. Comparative antioxidant activity of processed mango (*Mangifera indica*) and bush mango (*Irvingia gabonensis*) kernels. *Nigerian Food J.*, 30(2): 12-21.
- Atawodi SE, Bulus T, Ibrahim S, Ameh DA, Nok, AJ, MammN M & Galadima M 2003. *In vitro* trypanocidal effect of methanol extract of some Nigerian Savannah plants. *Afr. J. Biotech.*, 2(9): 312-321.
- Bridges CC & Zalups RK 2005. Molecular and ionic mimicry and the transport of toxicmetals. *Toxicol. Appl. Pharmacol.*, 204: 274-308.
- Burkill HM 2000. Useful Plants of West Tropical Africa. Second edition, volume 5, Royal Botani Garden Kewer, pp. 272-275.
- Cavanagh HM & Wilkinson JM 2002. Biological activities of lavender essential oil. *Phytother Res.* 16(4): 301-308.
- Chehregani A, Mohsenzadeh F, Mirazi N, Hajisadeghian S & Baghali Z 2010. Chemical composition and antibacterial activity of essential oils of Tripleurospermum disciforme in three developmental stages. *Pharm. Biol.*, 48: 1280–1284.
- Cline M, Taylor JE, Flores J, Bracken S, McCall S & Ceremugu ET 2008. Investigation of the anxiolytic effects of linalool, a lavender extract, in the male Sprague-dawley rat. *AANA J.*, 76(1): 1-6.
- Eybl V & Kotizová D 2010. Protective effect of manganese in cadmium induced hepatic oxidative damage, changes in cadmium distribution and trace element levels in mice. *Interdisc. Toxicol.*, 3(2): 68-72.
- Fernandes ES, Passos GF, Medeiros R, Cunha FM, Ferreira J, Campos MM, Pianowski LF & Calixto JB 2007. Antiinflammatory effects of compounds alpha humulene and (-)-trans caryophyllene isolated from the essential oil of *Cordia verbenacea. Eur. J. Pharmacol.*, 569: 228-236.
- Gholivand MB, Rahimi-Nasrabadi M, Batooli H & Ebrahimabadi AH 2010. Chemical composition and antioxidant activities of the essential oil and methanol extracts of *Psammogeton canescens*. *Food and Chem. Toxicol.*, 48: 24–28.
- GÜne§ FE 2013. Medicinal uses of squalene as a natural antioxidant. *J. Marmara Univ. Institute of Health Sci.*, 3(4): 220-225.
- Iwu MM 1993. Handbook of African Medicinal Plants. CRC Press, London.
- Iwueke AV, Nwodo OFC & Okoli CO 2006. Evaluation of the anti-inflammatory and analgesic activities of *Vitex doniana* leaves. *Afri. J. Biotech.*, 5(20): 1929-1935.
- Jamakala O & Rani AU 2014. Mitigating role of zinc and iron against cadmium induced toxicity in liver and kidney of male albino rat: a study with reference to metallothionein quantification. *Int. J. Pharm. Pharm. Sci.*, 6(9): 411-417.
- Katende AB 1995. Useful trees and shrubs for Uganda. Identification, propagation and management for agricultural and pastoral communities.Regional soil conservation Unit (RSCU), Swedish International Development Authority (SIDA). *Molecules*, 5: 731-735.
- Klopell FC, Lemos M, Sousa JP, Comunello E, Maistro EL, Bastos JK & de Andrade SF 2007. Nerolidol, an antiulcer constituent from the essential oil of *Baccharis dracunculifolia* DC (Asteraceae). Z *Naturforsch*, 62: 537–542.

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- Koudou J, Abena AA, Ngaissona P & Bessie re JM 2005. Chemical composition and pharmacological activity of essential oil of *Canarium schweinfurthii*. *Fitoterapia*, 76: 700-703.
- Kundu A, Saha S, Walia S, Shakil NA, Kumar J & Annapurna K 2013. Cadinene sesquiterpenes from *Eupatorium* adenophorum and their antifungal activity. J. Environ Sci. Health, 48: 516-522.
- Lee MT, Lin WC, Yu B & Lee TT 2017. Antioxidant capacity of phytochemicals and their potential effects on oxidative status in animals – A review. *Asian-Australas J. Anim. Sci.*, 30: 299-308.
- Legault J, Dahl W, Debiton E, Pichette A & Madelmont JC 2003. Antitumor activity of balsamfir oil: Production of reactive oxygen species induced by α-humulene as possible mechanism of action. *Planta. Med.*, 69: 402-407.
- Liu MC, Xu Y, Chen YM, Li J, Zhao F, Zheng C, Jing JF, Ke T, Chen JY & Luo WJ 2013. The effect of sodium selenite on lead induced cognitive dysfunction. *Neurotoxicol*, 36:82-88.
- Lopes NP, Massuo MJ, Andrade EHA, Maia JGS, Yoshida M, Planchart AR, & Katzin AM 1999. Antimalarial use of volatile oil from leaves of *Virola surinamensis*. J. Ethnopharmacol., 67: 313–319.
- Mahmoud GI 2013. Biological effects, antioxidant and anticancer activities of marigold and basil essential oils. *J. Med. Plants Res.* 7:10: 561 - 572.
- Maria SA, Edison O & Patricia CG 2016. Linalool reverses neuropathological and behavioural impairments in old triple transgenic Alzheimer's mice. *Neuropharmacol.*, 102: 111-120.
- Nnamani CV, Oselebe HO & Ogbonna AN 2007. Effects of leaf extracts of *Dracean aborae* L and *Vitex donian* Sweet on the larvae of *Anopheles* mosquito. *Anim. Res. Intl.*, 5(2): 184-186.
- Peana AT, de Montis MG, Nieddu E, Spano MT, D'Aquila PS & Pippia P 2004. Profile of spinal and supra-spinal antinociception of (-)-linalool. *Eur. J. Pharmacol.*, 485: 165-174.
- Pengelly A 2004. The Constituents of Medicinal Plants; Allen and Unwin.
- Petrovska BB 2012. Historical review of medicinal plants' usage. *Pharmacognosy Reviews*, 6: 1-5.
- Phaniendra A, Jestadi DB, & Periyasa L 2015. Free radicals: properties, sources, targets, and their implication in various diseases. *Ind. J. Clin. Biochem.*, 30(1): 11-26.
- Sarikurkcu C, Sabih Ozer M, Cakir A, Eskici M & Mete E 2013. GC/ MS evaluation and *in vitro* antioxidant activity of essential oil and solvent extracts of an endemic plant used as folk remedy in Turkey: *Phlomis bourgaei* Boiss. *Evid. Based Complement Alternat. Med.* 293080.
- Seol G, Kang P, Lee HS & Seol GH 2016. Antioxidant activity of linalool in patient with carpal tunnel syndrome. *BMC Neurology*, 16: 17-24.
- Sofowora AE 1993. The State of Medicinal Plants in Nigeria, University of Ibadan, Ibadan, Nigeria.

- Sperotto ARM, Moura DJ, Péres VF, Damasceno FC, Caramão EB, Henriques JAP & Saffi J 2013. Cytotoxic mechanism of *Piper gaudichaudiamum* Kunth essential oil and its major compound nerolidol. *Food and Chem. Toxicol.*, 57: 57-68.
- Sun X, Wang S, Li T & Yang Y 2015. Anticancer activity of linalool terpenoid: Apoptosis induction and cell cycle arrest in prostate cancer cells. *Trop. J. Pharm. Res.*, 14(4): 619-625.
- Taghreed AI 2012. Chemical composition and biological activity of extracts from *Salvia bicolar* Deaf. Growing in Egypt. *Molecules*, 11315-11334.
- Tarasub N, Tarasub C, Watcharaporn D & Ayutthaya WDN 2011. Protective role of curcumin on cadmium-induced nephrotoxicity in rats. J. Environ. Chem. Ecotoxicol., 3(2): 17-24.
- Vinholes J, Rudnitskaya A, Goncalves P, Martel F, Coimbra MA & Rocha SM 2013. Hepatoprotection of sesquiterpenoids; a quantitative structure-activity relationship (QSAR) approach. *Food Chem.*, 146: 78 – 84.
- Vinholes J, Gonçalves P, Martel F, Coimbra MA & Rocha SM 2014. Assessment of the antioxidant and antiproliferative effects of sesquiterpenic compounds in in vitro Caco-2 cell models. *Food Chem.*, 156: 204–211.
- Wang C, Li CJ, Yang JZ, Ma J, Chen XG, Hou Q & Zhang DM 2013. Anti-inflammatory sesquiterpene derivatives from the leaves of *Tripterygium wilfordii*. J. Natural Prod., 76: 85-90.
- Wang C, Yoon SH, Jang HI, Chung YR, Kim JY, Choi ES & Kim SW 2011. Metabolic engineering of *Escherichia coli* for α-farnesene production. *Metabolic Engineering*, 13: 648-655.
- Wang DC, Sun SH, Shi LN, Qui DR, Li X, Wei DS, Zhang YM & Qin JC 2015. Chemical composition, antibacterial and antioxidant activity of the essential oils of *Metaplexis japonica* and their antibacterial components. *Int. J. Food Sci. and Techn.*, 50: 449-457.
- Wang J, Zhao J, Liu H, Zhou L, Liu Z, Wang J, Han J, Yu Z & Yang F 2010. Chemical analysis and biological activity of the essential oils of two Valerianaceous species from China: Nardosstachys chinensis and Valeriana officinalis. Molecules, 15: 6411-6422.
- Wu Q, Yu L, Qiu J, Shen B, Wang D, Soromou LW & Feng H 2014. Linalool attenuates lung ginflammation induced by Pasteurella multocida via activating Nrf-2 signaling pathway. *Int. Immunopharmacol.*, 21(2): 456–63.
- Yakubu OE, Ojogbane E, Nwodo OFC, Nwaneri-Chidozie VO & Dasofunjo K 2013. Comparative antioxidant and hypoglycaemic effects of aqueous, ethanol and n hexane extracts of leaf of *Vitex doniana*on streptozotocin induced diabetes in albino rats. *Afri. J. Biotech.*, 12(40): 5933-5940.
- Zhai Q, Narbad A & Chen W 2015. Dietary strategies for the treatment of cadmium and lead oxicity. *Nutrients*, 7: 552-555.